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# Complete Genome Sequence of *Enterobacter cloacae* B2-DHA, a Chromium-Resistant Bacterium

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Previously, we reported a chromium-resistant bacterium, *Enterobacter cloacae* B2-DHA, isolated from the landfills of tannery industries in Bangladesh. Here, we investigated its genetic composition using massively parallel sequencing and comparative analysis with other known *Enterobacter* genomes. Assembly of the sequencing reads revealed a genome of ~4.19 Mb in size.

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The chromium-resistant strain B2-DHA was isolated from the landfills of leather manufacturing tannery industries in the Hazaribagh area, in very close vicinity of the capital city Dhaka, Bangladesh, where the tannery wastes have been disposed for many years (1). Sequencing of the genomic DNA of B2-DHA was performed by an Illumina sequencer HiSeq-2500 PE106 (106-bp paired-end) with a single sequencing index. Read quality checks were performed with FastQC (2) version 0.10.1. Adapter and quality trimming on raw reads were conducted with Cutadapt (3). k-mer error correction was performed on the adapter-free reads using Quake version 0.3.5 (4). Properly paired reads were extracted from the corrected read pool, and the remaining singleton reads were listed as single-end reads. Both corrected paired-end and single-end reads are used in the subsequent *de novo* assembly. SOAPdenovo (5) version 2.04 was utilized to perform *de novo* assembly optimization with the error-corrected reads. A wide range of k-mers (29 to 99) was tried to identify the scaffold sequences with the maximal  $N_{50}$ . The largest  $N_{50}$ , 492,970 bp, was produced at the k-mer 97.

A total of 1,756,877,072 bases and 16,574,312 pairs of reads were generated by Illumina deep sequencing. Analysis of the raw reads with FastQC showed that the average per-base Phred score was  $\geq 36$  for all positions, and the mean per-sequence Phred score was 36. The overall G+C content was 55%. After quality trimming, error correction, and removal of the TruSeq adaptor sequence, 15,708,650 read pairs (94.78%) and 331,106 single-end sequences remained for further analysis. The set of scaffold sequences with maximal  $N_{50}$  (492,970 bp) was produced at a k-mer of 97. The corresponding scaffold sequences were subjected to gap closure using the corrected paired-end reads, and the resulting scaffolds ( $\geq 24,300$  bp) were defined as the final assembly. The final assembly was 4,218,945 bp and consisted of 13 scaffolds ranging from 72,208 bp to 777,700 bp.

The assembled genome sequence was annotated with the RAST (6) and Blast2GO (7) pipelines. ARAGORN (8) version 1.2.36 was used to predict tRNA genes. Prediction of tRNA-, rRNA- and protein-coding genes was performed based on RAST-predicted RNA genes. RAST resulted in 22 rRNA genes, including four long

subunit (LSU), 4 short subunit (SSU), eight 16S, and six 23S genes. GeneMark (9) and FGenesB (10) algorithms were applied, yielding 3,764 and 3,955 genes, respectively. A total of 3,955 protein-coding genes were predicted using FGenesB, of which 3,159 could be annotated by the Blast2GO pipeline. The functional annotation by RAST and Blast2GO indicated that B2-DHA contains many genes that are responsive to binding metal ions, like chromium, cobalt, copper, iron, arsenic, nickel, manganese, zinc, and potassium. For functional annotation, all protein-coding sequences resulting from GeneMark were used by Blast2GO. Based on the phylogenetic trees inferred by using the neighbor-joining method (11) presented in the MEGA6 software (12), B2-DHA resembles *Enterobacter cloacae* KMBC1 and *E. cloacae* EC7.

In summary, the strain B2-DHA harbors several metal-responsive genes that might be utilized in the bioremediation of chromium and other toxic metals in polluted environments.

**Nucleotide sequence accession numbers.** The genome sequence of B2-DHA strain has been registered in GenBank under accession no. LFJA00000000. The version described in this paper is the first version, LFJA00000000.1.

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## REFERENCES

1. Rahman A, Nahar N, Nawani NN, Jass J, Hossain K, Saud ZA, Saha AK, Ghosh S, Olsson B, Mandal A. 2015. Bioremediation of hexavalent chromium (VI) by a soil-borne bacterium, *Enterobacter cloacae* B2-DHA. *J Environ Sci Health, Part A* 50:1136–1147. <http://dx.doi.org/10.1080/10934529.2015.1047670>.
2. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc>.
3. Martin M. 2011. Cutadapt removes adapter sequences from high-

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- throughput sequencing reads. *EMBnet.J* 17:10–12. <http://dx.doi.org/10.14806/ej.17.1.200>.
4. Kelley DR, Schatz MC, Salzberg SL. 2010. Quake: quality-aware detection and correction of sequencing errors. *Genome Biol* 11:R116. <http://dx.doi.org/10.1186/gb-2010-11-11-r116>.
  5. Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, Tang J, Wu G, Zhang H, Shi Y, Liu Y, Yu C, Wang B, Lu Y, Han C, Cheung DW, Yiu SM, Peng S, Xiaoqian Z, Liu G, Liao X, Li Y, Yang H, Wang J, Lam TW, Wang J. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read *de novo* assembler. *GigaScience* 1:18. <http://dx.doi.org/10.1186/2047-217X-1-18>.
  6. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
  7. Götz S, García-Gómez JM, Terol J, Williams TD, Nagaraj SH, Nueda MJ, Robles M, Talón M, Dopazo J, Conesa A. 2008. High-throughput functional annotation and data mining with the Blast2GO suite. *Nucleic Acids Res* 36:3420–3435. <http://dx.doi.org/10.1093/nar/gkn176>.
  8. Laslett D, Canback B. 2004. ARAGORN, a program for the detection of transfer RNA and transfer-messenger RNA genes in nucleotide sequences. *Nucleic Acids Res* 32:11–16. <http://dx.doi.org/10.1093/nar/gkh152>.
  9. Borodovsky M, McIninch J. 1993. GENMARK: parallel gene recognition for both DNA strands. *Comput Chem* 17:123–133. [http://dx.doi.org/10.1016/0097-8485\(93\)85004-V](http://dx.doi.org/10.1016/0097-8485(93)85004-V).
  10. Salamov AA, Solovyev VV. 2000. *Ab initio* gene finding in *Drosophila* genomic DNA. *Genome Res* 10:516–522. <http://dx.doi.org/10.1101/gr.104.516>.
  11. Saitou N, Nei M. 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4:406–425. [PubMed](http://pubmed.ncbi.nlm.nih.gov/284322/).
  12. Tamura K, Stecher G, Peterson D, Filipiński A, Kumar S. 2013. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol Biol Evol* 30:2725–2729. <http://dx.doi.org/10.1093/molbev/mst197>.